



Modelling biochemical signalling pathways

or.. computing science meets the life sciences

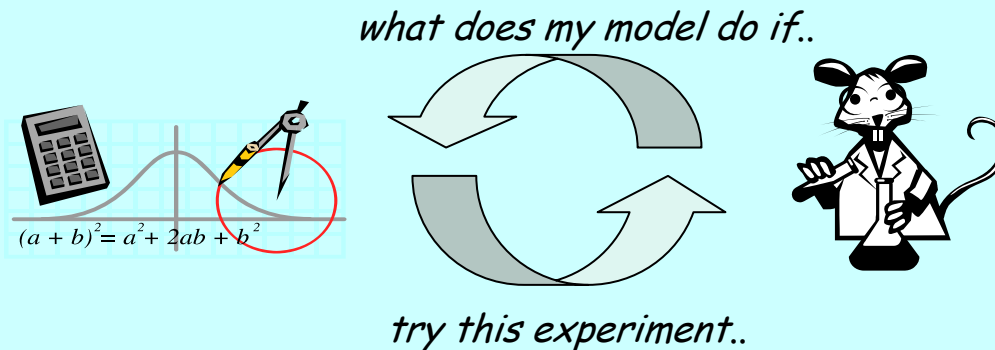
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Joint work with
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Vlad Vyshemirsky, Glasgow University

2007

The hypothesis

- Computing science can inspire the way we model and reason about biological systems, particularly ones involving communication
- Not biologically inspired computing, but **computationally inspired biology/biochemistry**
- Modelling for intervention (e.g. drug targets)

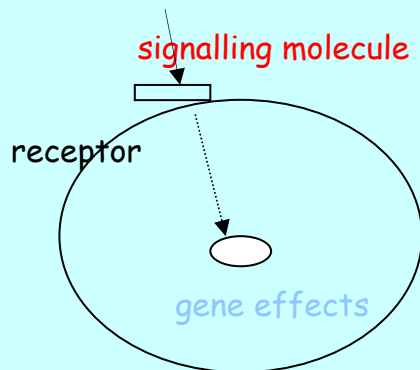


The talk

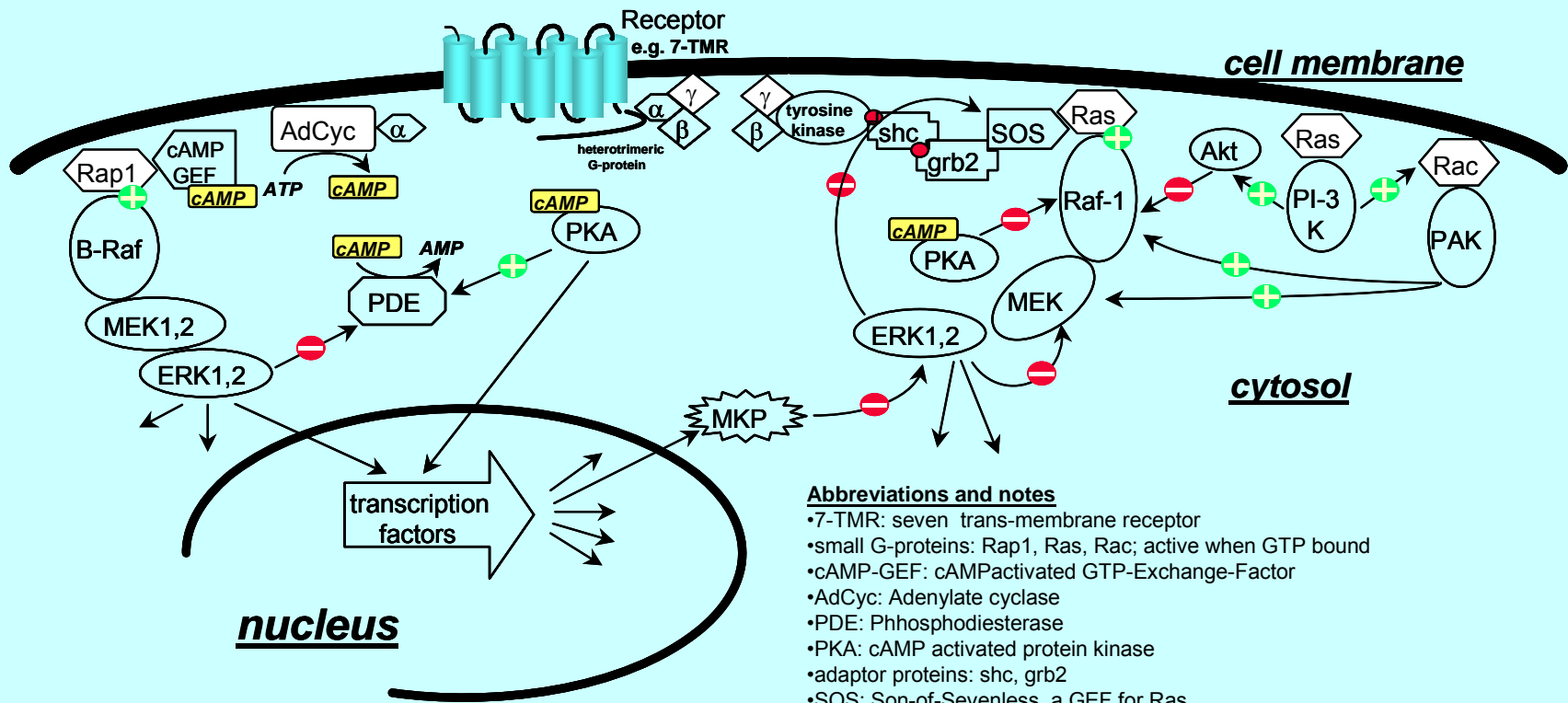
- gentle/naive biochemistry - signalling pathways
- new models of dynamic behaviour of signalling pathways based on CTMCs (continuous time Markov chains)
 - stochastic process algebra (PEPA)
 - model checking continuous stochastic logic (PRISM)
- new quantitative analysis
- relation to traditional ODE models
- Lots of Acronyms
 - RKIP/ERK
 - PEPA
 - ODE
 - PRISM
 - DIZZY

Cell signalling for dummies

- movement of signal from outside cell to nucleus
- fundamental to cell processes (growth, division, differentiation, apoptosis)
- signalling is via membrane receptors, "signal" is *phosphorylation - accumulation of certain phosphorylated proteins*



A little more complex.. pathways/networks

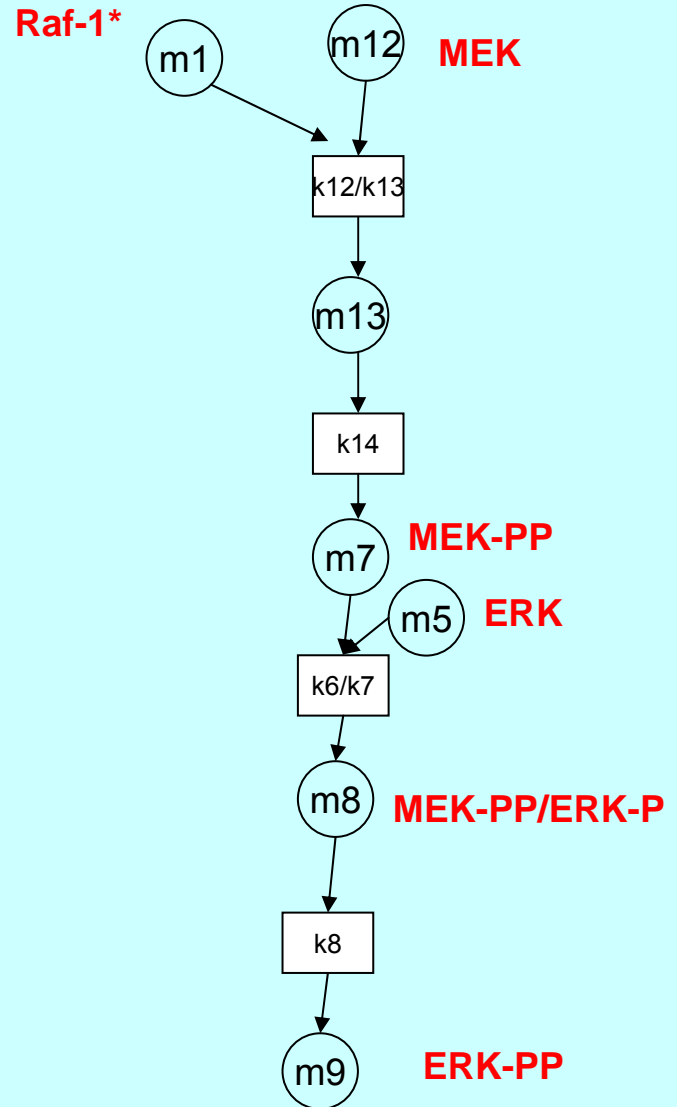
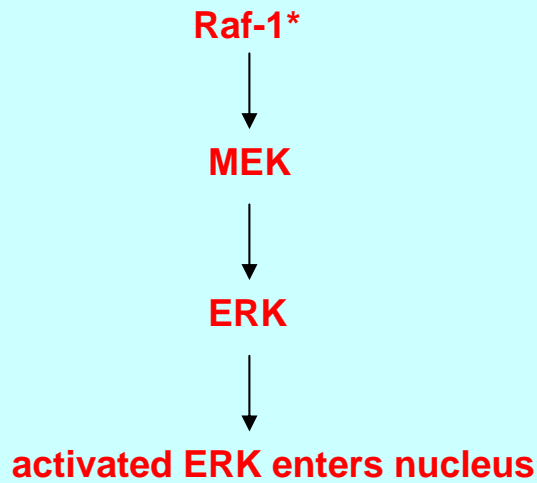


Abbreviations and notes

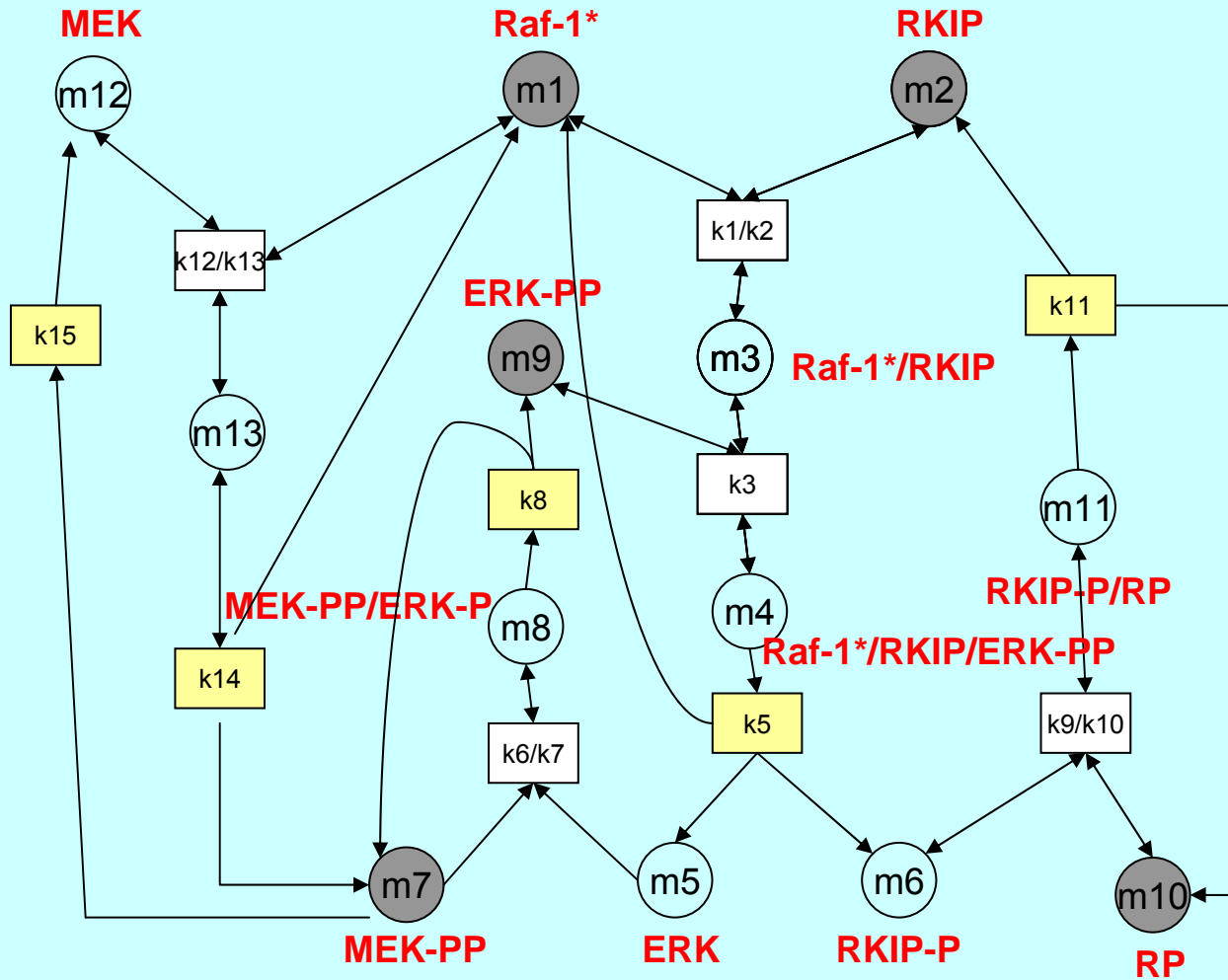
- 7-TMR: seven trans-membrane receptor
- small G-proteins: Rap1, Ras, Rac; active when GTP bound
- cAMP-GEF: cAMPactivated GTP-Exchange-Factor
- AdCyc: Adenylate cyclase
- PDE: Phosphodiesterase
- PKA: cAMP activated protein kinase
- adaptor proteins: shc, grb2
- SOS: Son-of-Sevenless, a GEF for Ras
- PI-3 K: Phosphatidylinositol-3 kinase
- Akt: a kinase activated by PI-3 K via PI-3 and another kinase, PDK
- PAK: a kinase activated by binding to Rac
- MKP: MAPK phosphatase, dephosphorylates MAPKs

+ activation
 - inhibition
 • phosphorylation

Basic ERK Pathway

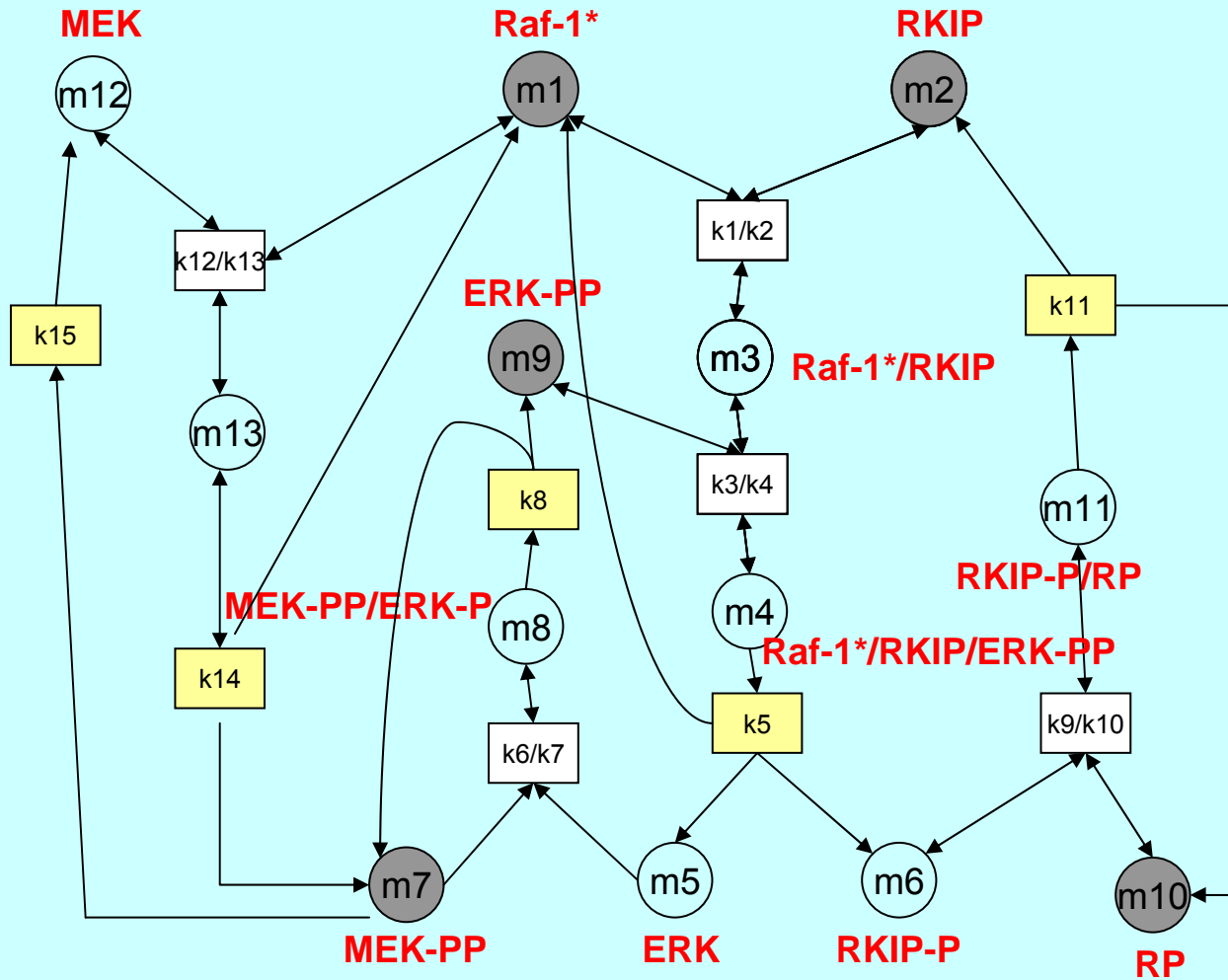


RKIP Inhibited ERK Pathway



CANCER RESEARCH UK
BEATSON LABORATORIES

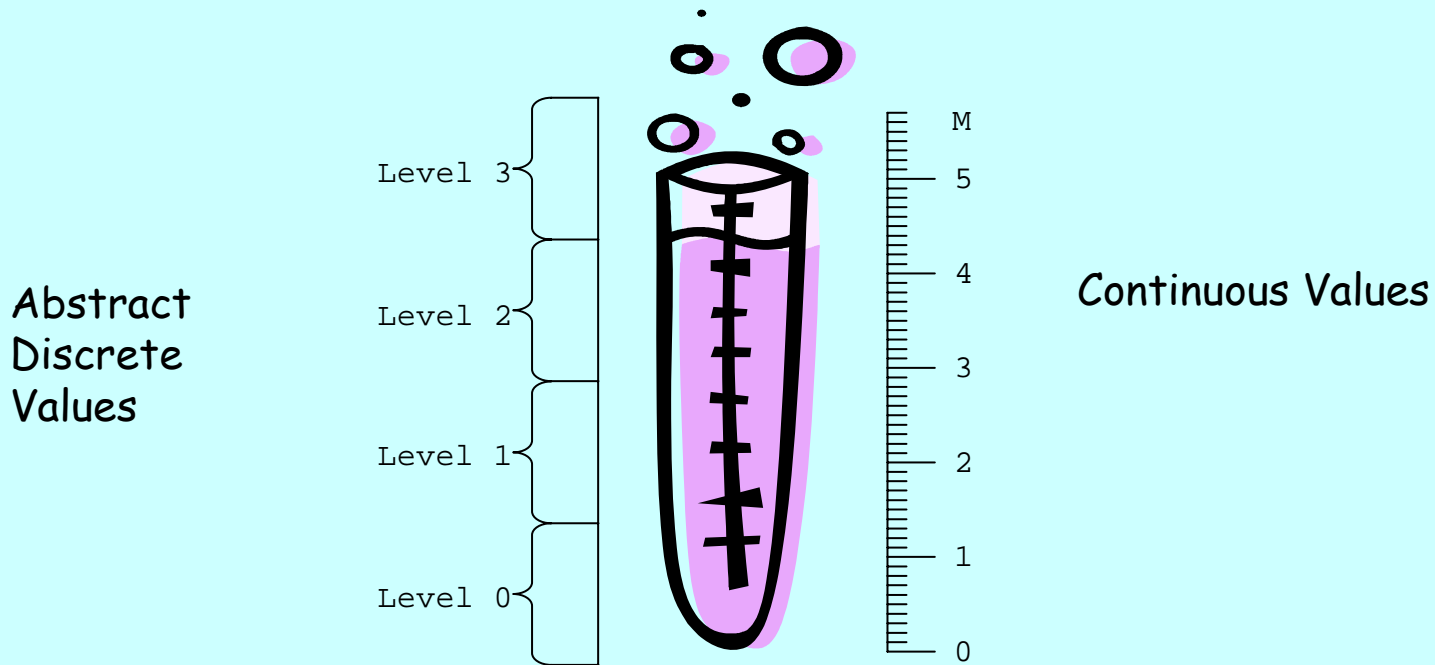
RKIP Inhibited ERK Pathway



producer/consumer behaviour

- computation
- stochastic
- concurrent
- message passing

Modelling *concentrations* not molecules



levels 0..N represent $[0, 1 \cdot M/N)$, $[1 \cdot M/N, 2 \cdot M/N)$, ... , $[N-1 \cdot M/N, N \cdot M/N]$

nb. time is **real**, concentration is **discrete**.

PEPA

Process algebra with *performance*, invented by Jane Hillston

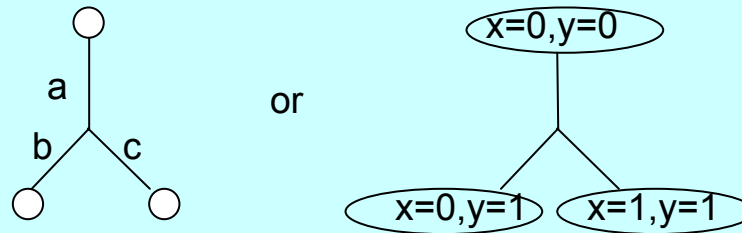
<i>Prefix</i>	(a,r)	
<i>Choice</i>	$P1 + P2$	competition between components (race)
<i>Cooperation/</i>	$P1 P1$	$a \notin l$ independent concurrent (interleaved) actions
<i>Synchronisation</i>		$a \in l$ shared action, <u>at rate of slowest</u> <i>multiway</i>
<i>Constant</i>	$A = P$	assign names to components

$$P ::= S \mid P \mid\mid P$$
$$S ::= (\alpha,r).S \mid S+S \mid A$$

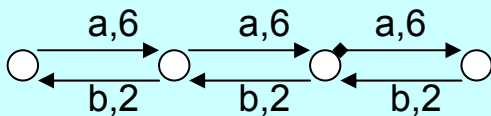
Synchronisation is associative, commutative.
T is the passive rate.

Process algebra semantics

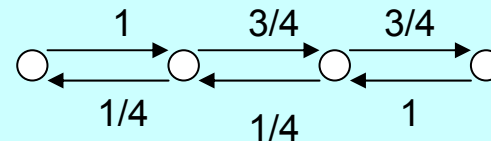
Standard process algebra descriptions denote a form of labelled transition systems or Kripke structures.



Performance evaluation process algebra descriptions denote Continuous Time Markov chains (probabilistic transition systems).

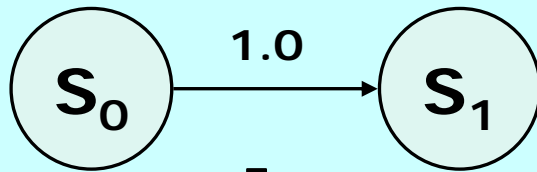


CTMC

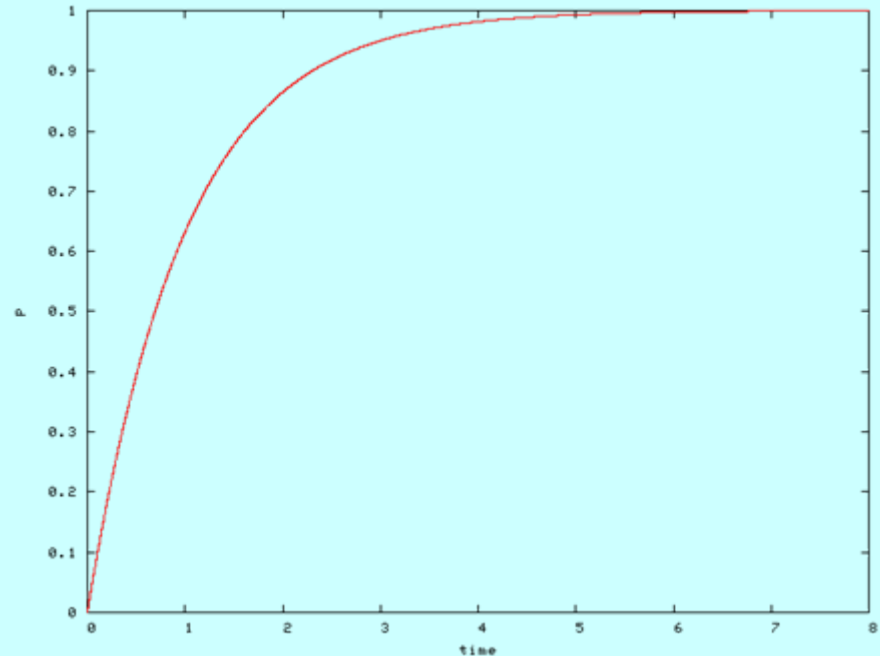


DTMC

Continuous Time Markov Chains



$$P(t) = 1 - e^{-\lambda t}$$
$$\lambda = 1.0$$



State based models for *dynamic, stochastic* behaviour.

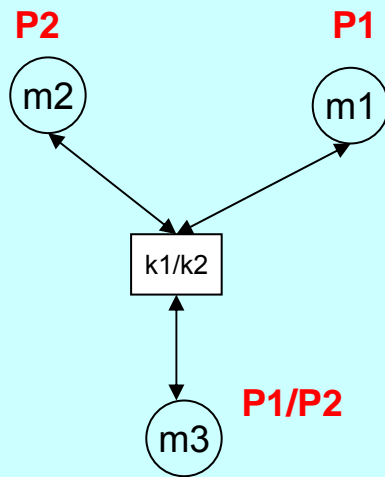
- performance coefficients associated with all transitions
- from rates we derive probabilities
- "memoryless" property.

Modelling the ERK Pathway in PEPA

- Each *reaction* is modelled by an event, which has a *performance coefficient*.
- Each *protein* is modelled by a process which synchronises others involved in a reaction. A fine-grained distributed view.
(*reagent-centric view*)
- Each *sub-pathway* is modelled by a process which synchronises with other sub-pathways. A coarser grained view.
(*pathway-centric view*)

Take the simplest concentration abstraction, $N=1$, the "high/low" approach.

Modelling reactions



Reaction	Producer(s)	Consumer(s)
$k1react$	$\{P2,P1\}$	$\{P1/P2\}$
$k2react$	$\{P1/P2\}$	$\{P2,P1\}$

$k1react$ and $k2react$ will be a 3-way synchronisations.

(Multiway synch is essential!)

Modelling reactions

Reagent view: models whether or not a reagent can participate in a reaction (observable/unobservable).

Each reagent gives rise to a pair of definitions.

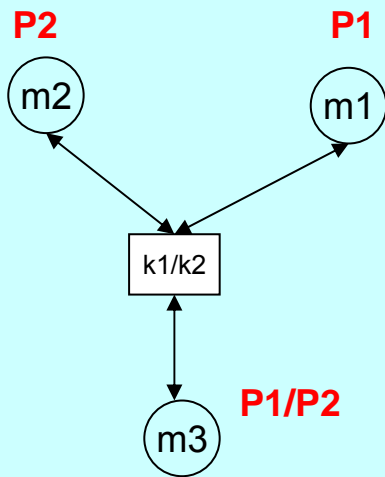
Model equations

$$P1_H = (k1react, k1). P1_L$$
$$P1_L = (k2react, T). P1_H$$

$$P2_H = (k1react, k1). P2_L$$
$$P2_L = (k2react, T). P2_H$$

$$P1/P2_H = (k2react, k2). P1/P2_L$$
$$P1/P2_L = (k1react, T). P1/P2_H$$

(consumers have passive rate)



Modelling reactions

Reagent view: models whether or not a reagent can participate in a reaction (observable/unobservable).

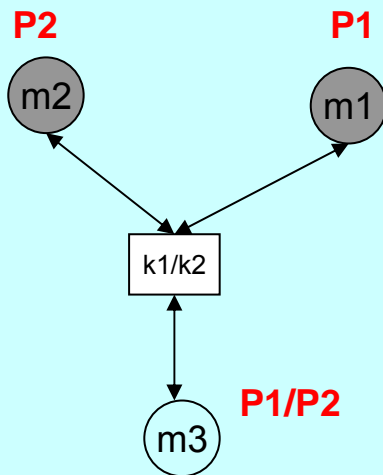
Each reagent gives rise to a pair of definitions.

Model equations

$$P1_H = (k1react, k1). P1_L$$
$$P1_L = (k2react, T). P1_H$$

$$P2_H = (k1react, k1). P2_L$$
$$P2_L = (k2react, T). P2_H$$

$$P1/P2_H = (k2react, k2). P1/P2_L$$
$$P1/P2_L = (k1react, T). P1/P2_H$$



Model configuration

$$P1_H \mid k1react, k2react \mid P1_H \mid k1react, k2react \mid P1/P2_L$$

(assuming initial concentrations of m1 and m2)

Modelling reactions

N levels:

Model equations

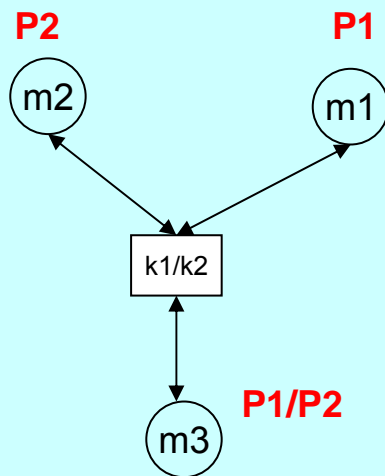
$$P1_N = (k1react,N * k1). P1_{N-1}$$
$$P1_{N-1} = (k1react,(N-1) * k1). P1_{N-2} + (k2react,T). P1_N$$

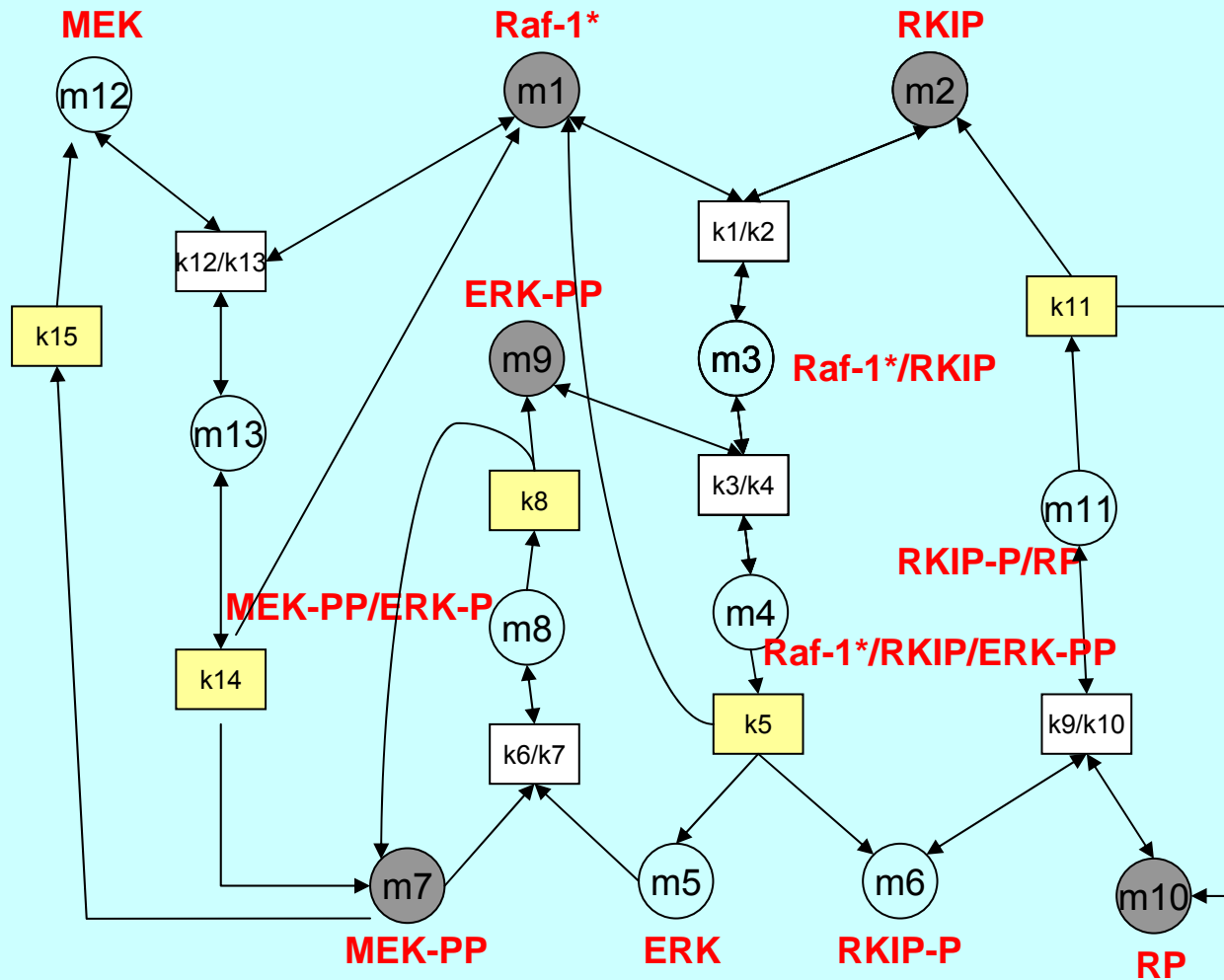
...

$$P1_1 = (k1react,k1). P1_0 + (k2react,T). P1_2$$

$$P1_0 = (k2react,T). P1_1$$

....





Reagent view:

$Raf-1^*_H = (k1react, k1)$. $Raf-1^*_L + (k12react, k12)$. $Raf-1^*_L$

$Raf-1^*_L = (k5product, T)$. $Raf-1^*_H + (k2react, T)$. $Raf-1^*_H + (k13react, T)$. $Raf-1^*_H + (k14product, T)$. $Raf-1^*_H$

...

(26 equations)

Signalling Dynamics

Reagent view: model configuration

Raf-1*_H | k1react,k12react,k13react,k5product,k14product |

RKIP_H | k1react,k2react,k11product |

Raf-1*_H/RKIP_L | k3react,k4react |

Raf-1*/RKIP/ERK-PP_L | k3react,k4react,k5product |

ERK-P_L | k5product,k6react,k7react |

RKIP-P_L | k9react,k10react |

RKIP-P/RP_L | k9react,k10react,k11product |

RP_H ||

MEK_L | k12react,k13react,k15product |

MEK/Raf-1*_L | k14product |

MEK-PP_H | k8product,k6react,k7react |

MEK-PP/ERK_L | k8product |

MEK-PP_H | k8product |

ERK-PP_H

Modelling reactions

Pathway view: model *chains* of behaviour flow

Model equations

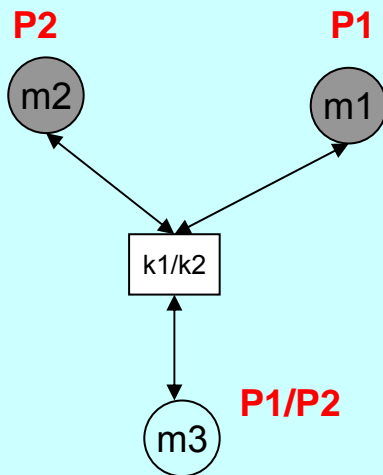
$\text{Pathway}_1 = (k1_{\text{react}}, k1). (k2_{\text{react}}, k2). \text{Pathway}_1$

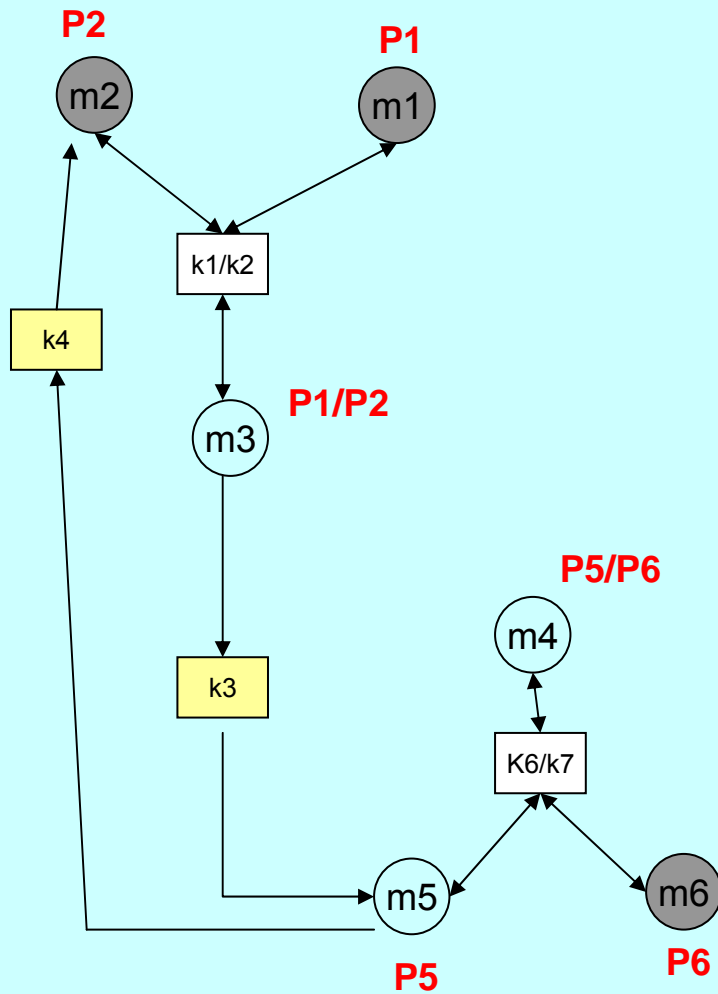
Model configuration

Pathway_1

(assuming initial concentrations of m1 and m2)

Note: only one component!





Pathway view: model chains of behaviour flow.

Two pathways, corresponding to initial concentrations:

$$\text{Path10} = (\text{k1react}, \text{k1}). \text{Path11}$$

$$\text{Path11} = (\text{k2react}). \text{Path10} + (\text{k3product}, \text{k3}). \text{Path12}$$

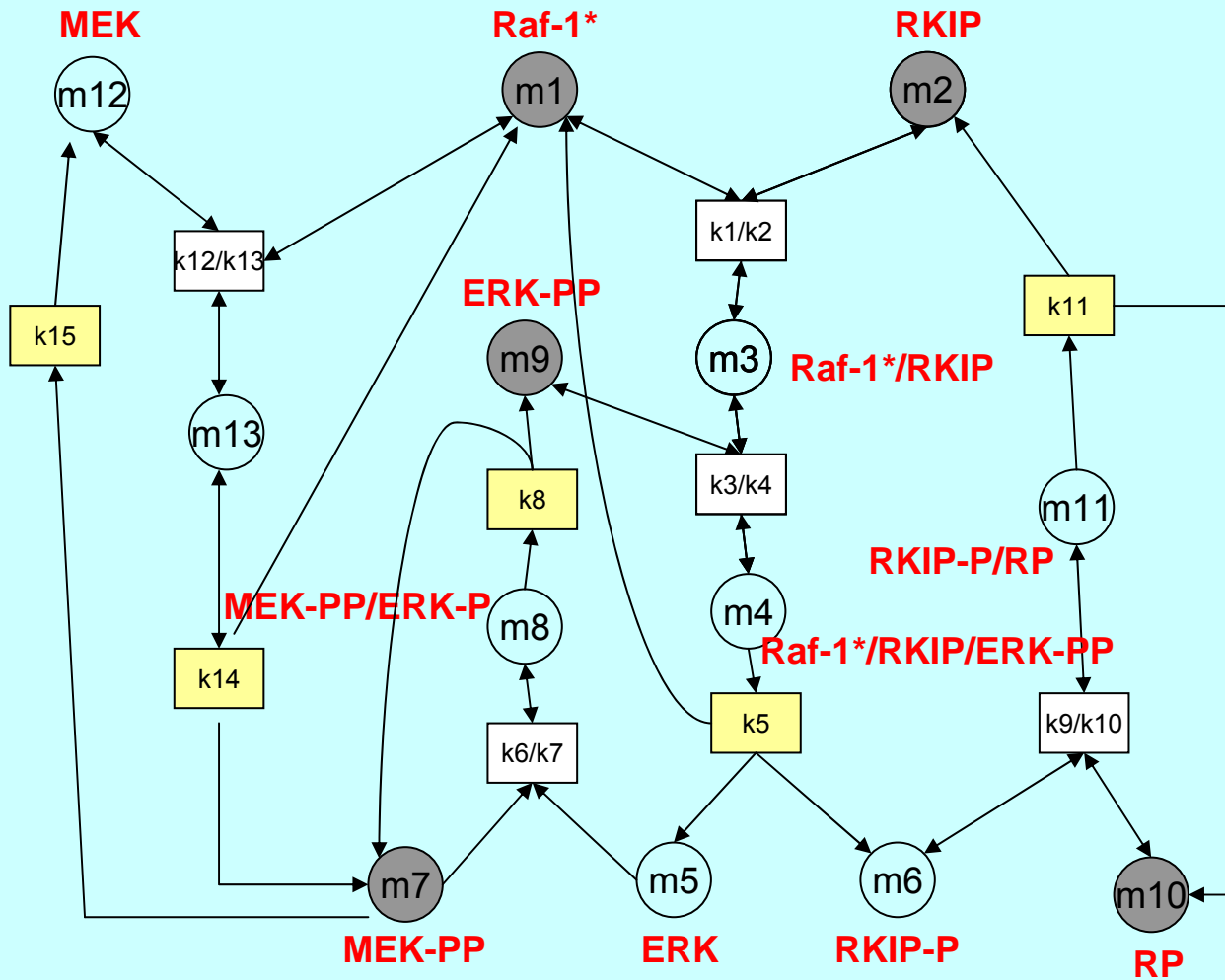
$$\text{Path12} = (\text{k4product}, \text{k4}). \text{Path10} + (\text{k6react}, \text{k6}). \text{Path13}$$

$$\text{Path13} = (\text{k7react}, \text{k7}). \text{Path12}$$

$$\text{Path2} = (\text{k6react}, \text{k6}). (\text{k7react}, \text{k6}). \text{Path2}$$

Pathway view: model configuration

$$\text{Path10} \mid \text{k6react}, \text{k7react} \mid \text{Path2}$$



Pathway view:

Pathway10 = (k9react,k9). ((k11product,k11). Pathway10 + (k10react,k10). Pathway10)

...

(5 pathways)

Pathway view: model configuration

Pathway10 |k12react,k13react,k14product| Pathway40

|k3react,k4react,k5product,k6react,k7react,k8product| Pathway30

|k1react,k2react,k3react,k4react,k5product| Pathway20

|k9react,k10react,k11product| Pathway50

simpler!

What is the difference?

- reagent-centric view is a *fine* grained view
- pathway-centric view is a *coarse* grained view
 - reagent-centric is easier to derive from data
 - pathway-centric allows one to build up networks from already known components

The two models are equivalent!

The equivalence proof, based on *bisimulation* between steady state solutions, unites two views of the same biochemical pathway.

Steady State Solution

1 0.04135079004156481
2 0.020806115102310632
3 0.07346775929692899
4 0.006935371700770152
5 0.06516104016641672
6 0.03737546622097119
7 0.011336715749471194
8 0.036048205933593286
9 0.004639841577167708
10 0.005691394350960237
11 0.04138456618620803
12 0.0025828089820320505
13 0.004807783620797024
14 0.04817123798507296
15 0.018640671069835055
16 0.016743539619515142
17 0.02162874351056745
18 0.0028912552492803816
19 0.004970238100423158
20 0.02076780718322302
21 0.1840054851485999
22 0.008846052672337585
23 0.01413218356459678
24 0.0030482221649047224
25 0.0020844704151460223
26 0.20477329233182312
27 0.09642576891046874
28 0.0012831731450123965

Reagent view

1 0.04135079004156353
2 0.020806115102310604
3 0.07346775929692419
4 0.006935371700769834
5 0.06516104016641262
6 0.03737546622096783
7 0.011336715749470889
8 0.03604820593359156
9 0.005691394350959787
10 0.004639841577167543
11 0.04138456618620752
12 0.04817123798507505
13 0.0025828089820318246
14 0.01864067106983504
15 0.004807783620796737
16 0.01674353961951507
17 0.020767807183224345
18 0.021628743510568222
19 0.18400548514860549
20 0.002891255249280038
21 0.008846052672337464
22 0.004970238100423424
23 0.014132183564597499
24 0.20477329233182964
25 0.09642576891047139
26 0.0030482221649046053
27 0.0020844704151453983
28 0.0012831731450119671

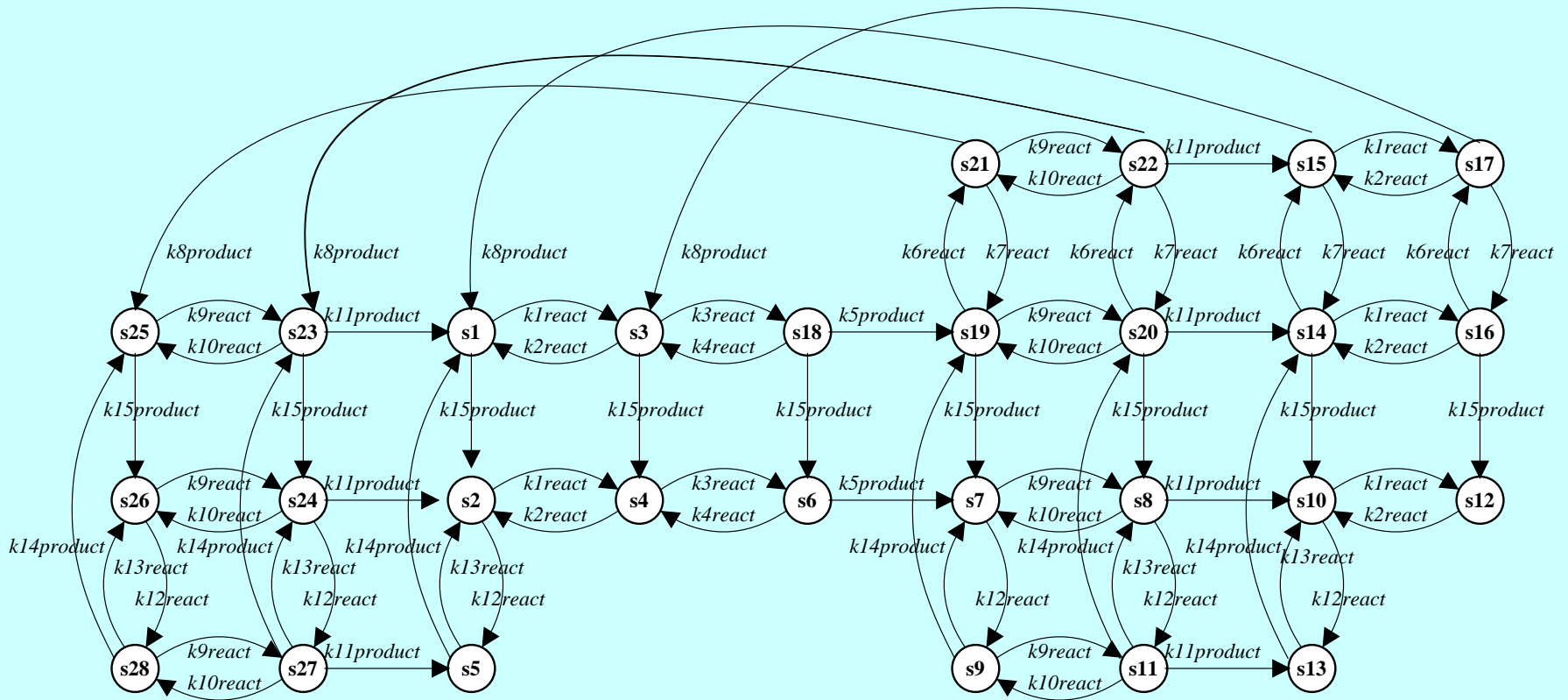
Pathway view

State space of reagent and pathway model

<p>state s1</p>	<p>reagent-view Raf-1*_H, RKIP_H, Raf-1*/RKIP_L, Raf-1*/RKIP/ERK-PP_L, ERK_L, RKIP-P_L, RKIP-P/RP_L, RP_H, MEK_L, MEK/Raf-1*_L, MEK-PP_H, MEK-PP/ERK_L, ERK-PP_H</p> <p>pathway view Pathway50, Pathway40, Pathway20, Pathway10</p>
<p>s2</p> <p>·</p> <p>·</p> <p>·</p> <p>s28</p>	<p>...</p>

(28 states)

State space of reagent and pathway model



What do you do with these two models?

-investigate properties of the underlying Markov model.

Transient analysis

e.g. analysis to determine whether a state will be reached.

Steady state analysis

e.g. analysis of the steady state solution - behaviour in the long run.

Note: there isn't one steady state, but a very large "cycle"!

Static Analysis

Check for deadlocks, livelocks

- something "wrong" with model
- e.g. an incompleteness (discovered problem with RKIP model)

Quantitative Analysis

Generate steady-state probability distribution (using linear algebra).

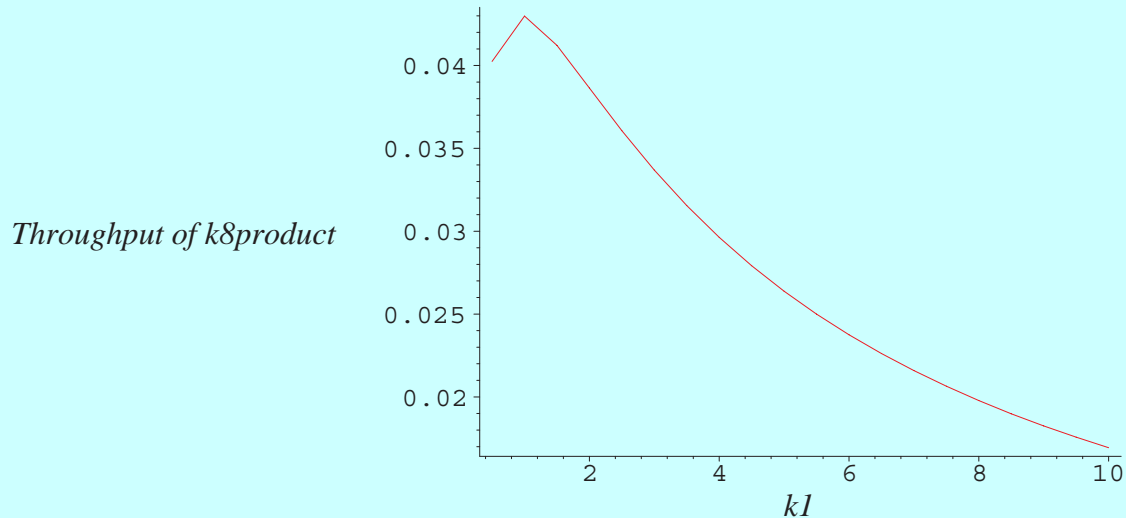
1. Use state finder (in reagent model) to aggregate probabilities and look for proteins high or low.

Example

increase k_1 from 1 to 100 and the probability of being in a state with ERK-PP_H drops from .257 to .005

2. Perform throughput analysis (in pathway model).

Quantitative Analysis



Effect of binding of RKIP to Raf-1* on ERK-PP

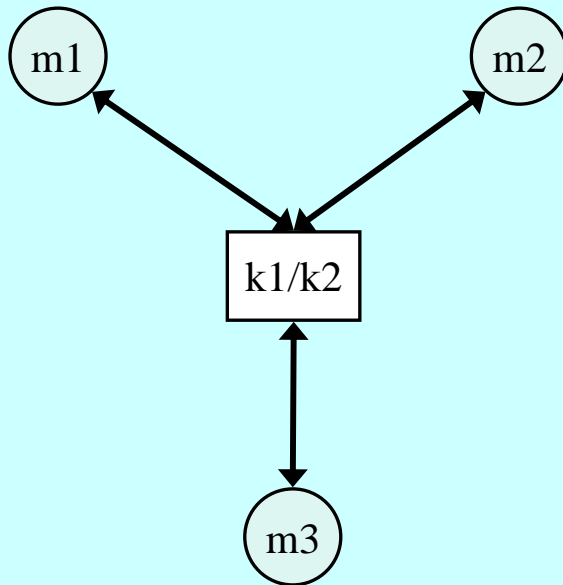
(increasing the rate of k_1 on k_8 product throughput (rate x probability))

We can see the effect of RKIP, but is this only indicative?

How accurate is the model?

Ordinary differential equations

Mass action kinetics (semantics):

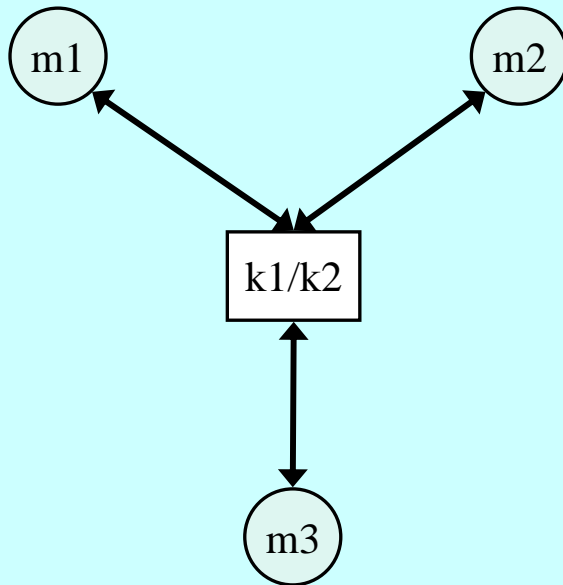


$$\left\{ \begin{array}{l} \frac{dm_1}{dt} = -k_1 m_1 m_2 + k_2 m_3 \\ \frac{dm_2}{dt} = -k_1 m_1 m_2 + k_2 m_3 \\ \frac{dm_3}{dt} = k_1 m_1 m_2 - k_2 m_3 \end{array} \right.$$

note, nonlinear ODEs (second order reactions)

more accurate rates

Mass action kinetics (semantics):



Consider forward only:

$$\frac{dm_3}{dt} = k_1 m_1 m_2$$

But the PEPA model has rate **k1**.
A consequence of synchronisation (slowest).

More accurate rates, and model checking capabilities will be obtained by moving to a state-based PRISM model.

PRISM reagent model

PRISM - probabilistic model checker
developed by Marta Kwiatkowska et al

Temporal reasoning (*experiments*, open formulae) for

- DTMCs
- MDPs
- CTMCs

- Prism modelling language - processes and synchronising transitions. Rates are *multiplied*.

```
const int N = 7;
const double R = 2.5/N /* 2.5 is initial concentration */
```

```
module RAF1
```

```
    RAF1: [0..N] init N;
    [r1] (RAF1 > 0) -> RAF1*R: (RAF1' = RAF1 - 1);
    [r2] (RAF1 < N) -> 1: (RAF1' = RAF1 + 1);
    [r5] (RAF1 < N) -> 1: (RAF1' = RAF1 + 1);
endmodule
```

```
module RKIP
```

```
    RKIP: [0..N] init N;
    [r1] (RKIP > 0) -> RKIP*R: (RKIP' = RKIP - 1);
    [r2] (RKIP < N) -> 1: (RKIP' = RKIP + 1);
    [r11] (RKIP < N) -> 1: (RKIP' = RKIP + 1);
endmodule
```

```
...
```

```
module Constants
```

```
    x: bool init true;
    [r1] (x) -> 0.53/R: (x' = true);
    [r2] (x) -> 0.0072/R: (x' = true);
    [r3] (x) -> 0.625/R: (x' = true);
    [r4] (x) -> 0.00245/R: (x' = true);
```

```
system RAF1 || RKIP || RAF1/RKIP ... || Constants endsystem
```

```
const int N = 7;
const double R = 2.5/N /* 2.5 is initial concentration */
```

```
module RAF1
```

```
  RAF1: [0..N] init N;
  [r1] (RAF1 > 0) -> RAF1*R: (RAF1' = RAF1 - 1);
  [r2] (RAF1 < N) -> 1: (RAF1' = RAF1 + 1);
  [r5] (RAF1 < N) -> 1: (RAF1' = RAF1 + 1);
endmodule
```

```
module RKIP
```

```
  RKIP: [0..N] init N;
  [r1] (RKIP > 0) -> RKIP*R: (RKIP' = RKIP - 1);
  [r2] (RKIP < N) -> 1: (RKIP' = RKIP + 1);
  [r11] (RKIP < N) -> 1: (RKIP' = RKIP + 1);
endmodule
```

```
...
```

```
module Constants
```

```
  x: bool init true;
  [r1] (x) -> 0.53/R: (x' = true);
  [r2] (x) -> 0.0072/R: (x' = true);
  [r3] (x) -> 0.625/R: (x' = true);
  [r4] (x) -> 0.00245/R: (x' = true);
```

```
system RAF1 || RKIP || RAF1/RKIP ... || Constants endsystem
```

rates

Example rate of r1: $(N \cdot R) \cdot (N \cdot R) \cdot 1 \cdot 0.53/R = 7 \cdot 0.53 \cdot 2.5$

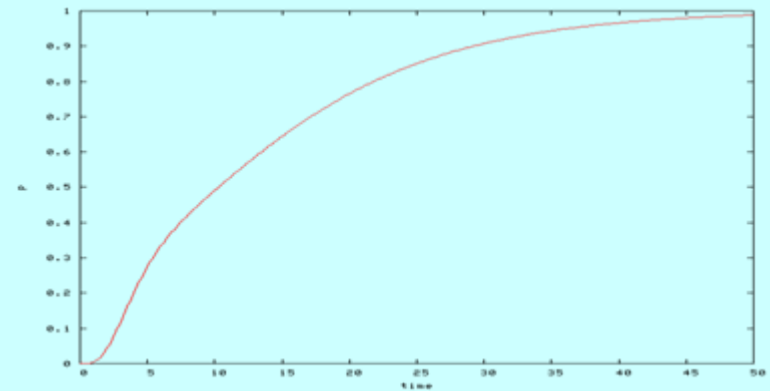
*RAF1 * RKIP * RAF1/RKIP * Constants*

Temporal properties

1. stability of protein (steady state)
2. monotonic decrease of protein (transient)
3. protein stability with varying rates (steady state)
4. protein activation order (transient)

CSL (Continuous Stochastic Logic)

Operator	CSL Syntax
True	$true$
False	$false$
Conjunction	$\phi \wedge \phi$
Disjunction	$\phi \vee \phi$
Negation	$\neg \phi$
Implication	$\phi \Rightarrow \phi$
Next	$P_{\bowtie p}[\mathbf{X}\phi]$
Unbounded Until	$P_{\bowtie p}[\phi \mathbf{U} \phi]$
Bounded Until	$P_{\bowtie p}[\phi \mathbf{U}^{\leq t} \phi]$
Bounded Until	$P_{\bowtie p}[\phi \mathbf{U}^{\geq t} \phi]$
Bounded Until	$P_{\bowtie p}[\phi \mathbf{U}^{[t_1, t_2]} \phi]$
Steady-State	$S_{\bowtie p}[\phi]$



$$P_{=?}[(true)U_{\leq 150}(protein = N)]$$

$$S_{=?}[(protein > 10)]$$

$$P_{\geq 1}[(protein1 < C)U(protein1 > C)]$$

Temporal properties

1. stability of protein (steady state)

$$S_{\rightarrow} [(RAF1 \geq C-1) \wedge (RAF1 \leq C+1)]$$

2. monotonic decrease of protein (transient)

$$P_{\rightarrow=1} [(true) \cup ((Protein = C) \wedge P_{\rightarrow=0.95} [X(Protein = C-1)])]$$

$$P_{\rightarrow} [(true) \cup^{k=120} (Protein > C)\{RAF1=C\}]$$

3. stability with varying rates (steady state)

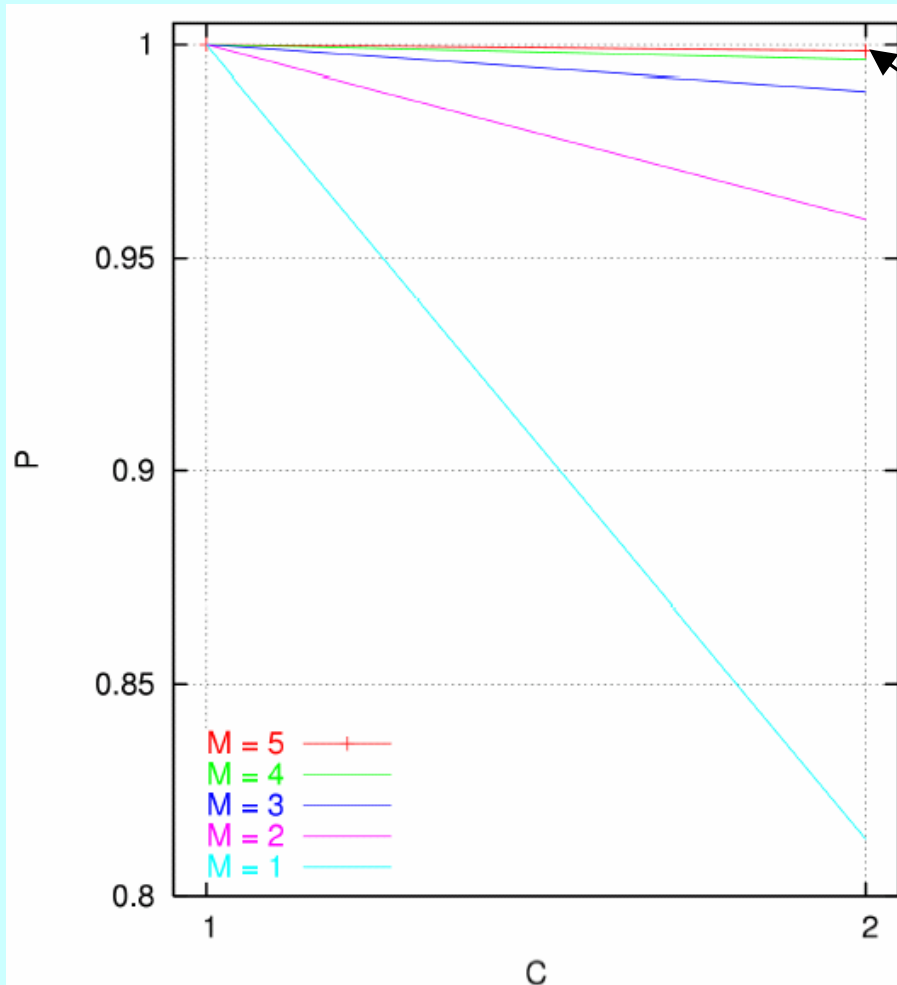
$$S_{\rightarrow} [(RAF1 \geq 2) \wedge (RAF1 \leq 3)]$$

$$S_{\rightarrow} [(RAF1 \geq 0) \wedge (RAF1 \leq 1)] \quad (\text{vary } k1)$$

4. protein activation order (transient)

$$P_{\rightarrow} [(RAF1/RKIP/ERK-PP < M) \cup (RAF1/RKIP = C)]$$

4. protein activation order



Probability that

RAF1/RKIP/ERK-PP reaches 5 before
 RAF1/RKIP reaches 2 > 99%

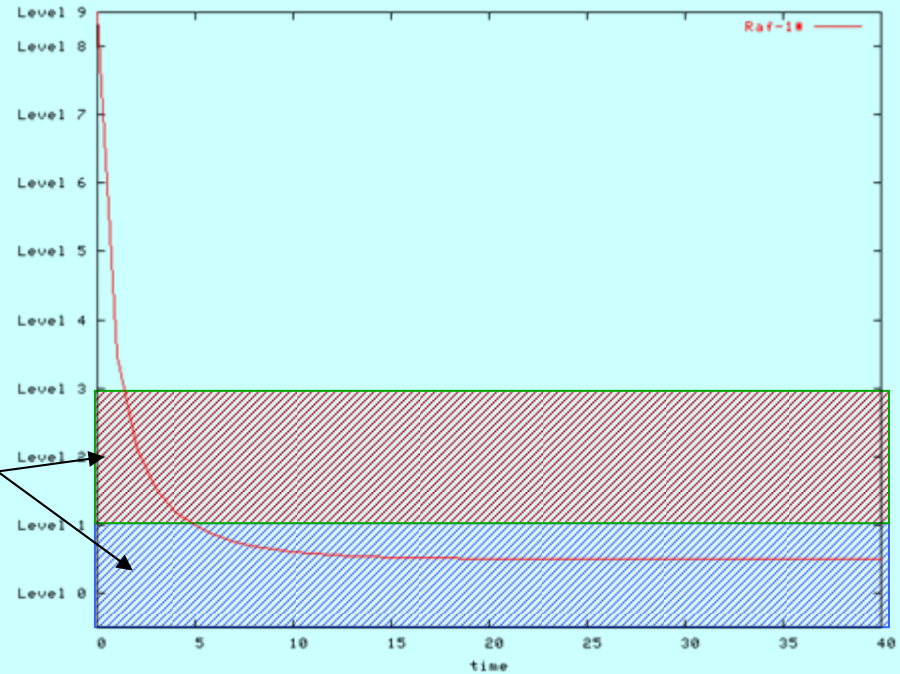
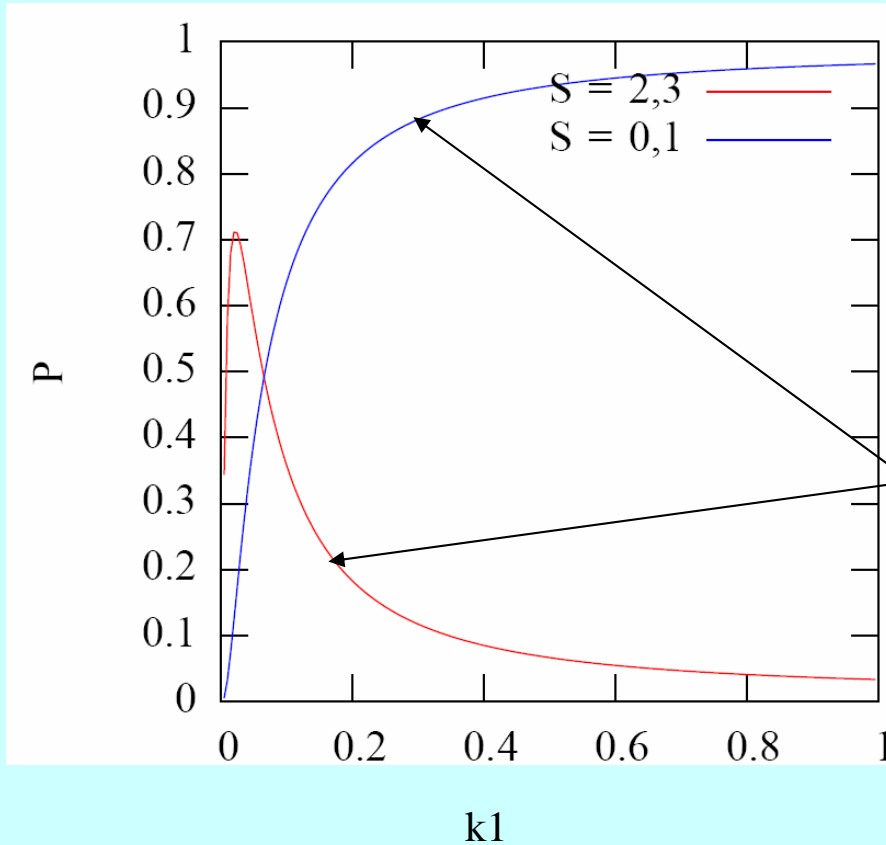
Property 4:

protein activation order (transient)

$P=?$

$[(RAF1/RKIP/ERK-PP < M) \cup (RAF1/RKIP = C)]$

3. Stability when varying rate



When the binding rate of RKIP and RAF1 (k_1) is increased, the probability that RAF1 stabilises on levels 2 or 3 (red square) falls, and the probability that RAF1 stabilises on levels 0 or 1 (blue square) rises.

Much more - allows us to conclude RKIP dampens down the ERK pathway.

Soundness

why is rate for r1 sound?

$$(RAF1^*R)^*(RKIP^*R)^*(k1^*N)$$

relate discrete and continuous variables: $m = m^*R = m/N$

Recall $\frac{dm_3}{dt} = k_1 m_1 m_2$

$m_3' = m_3 + (k1^* m_1^* m_2^* \Delta t)$. But abstract levels increase by 1, so

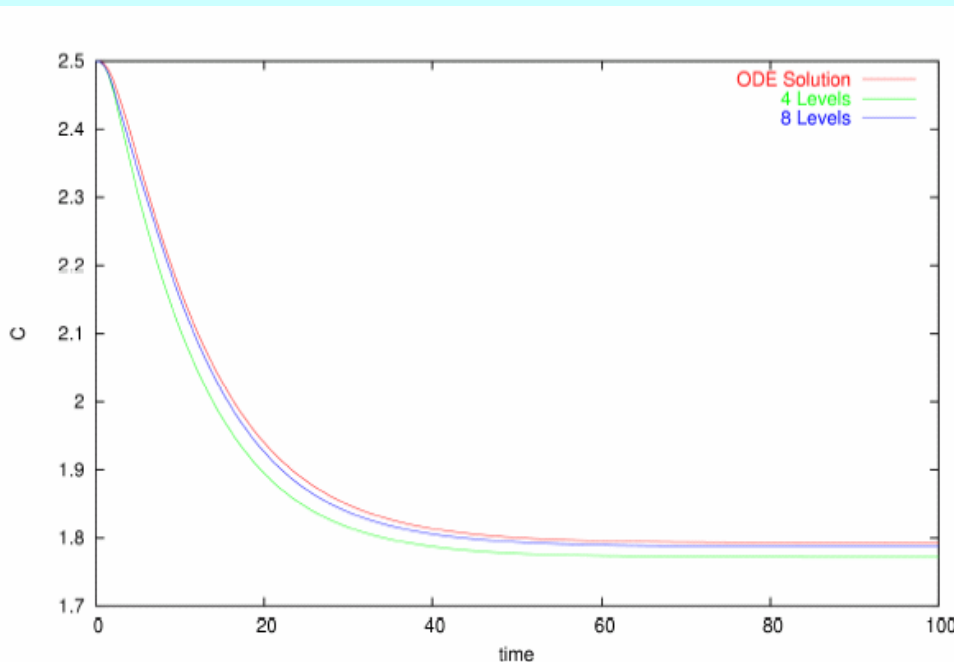
$$\Delta t = 1/(k1^* m_1^* m_2^* N)$$

$$\begin{aligned} \text{Let } \lambda &= k^* m_1^* m_2^* N \\ &= k^* (m_1^*R) (m_2^*R)^*N \\ &= k1^* (RAF1^*R)^*(RKIP^*R)^*N \end{aligned}$$

(We have, in effect, encoded Euler's method of numerical integration!)

Simulation

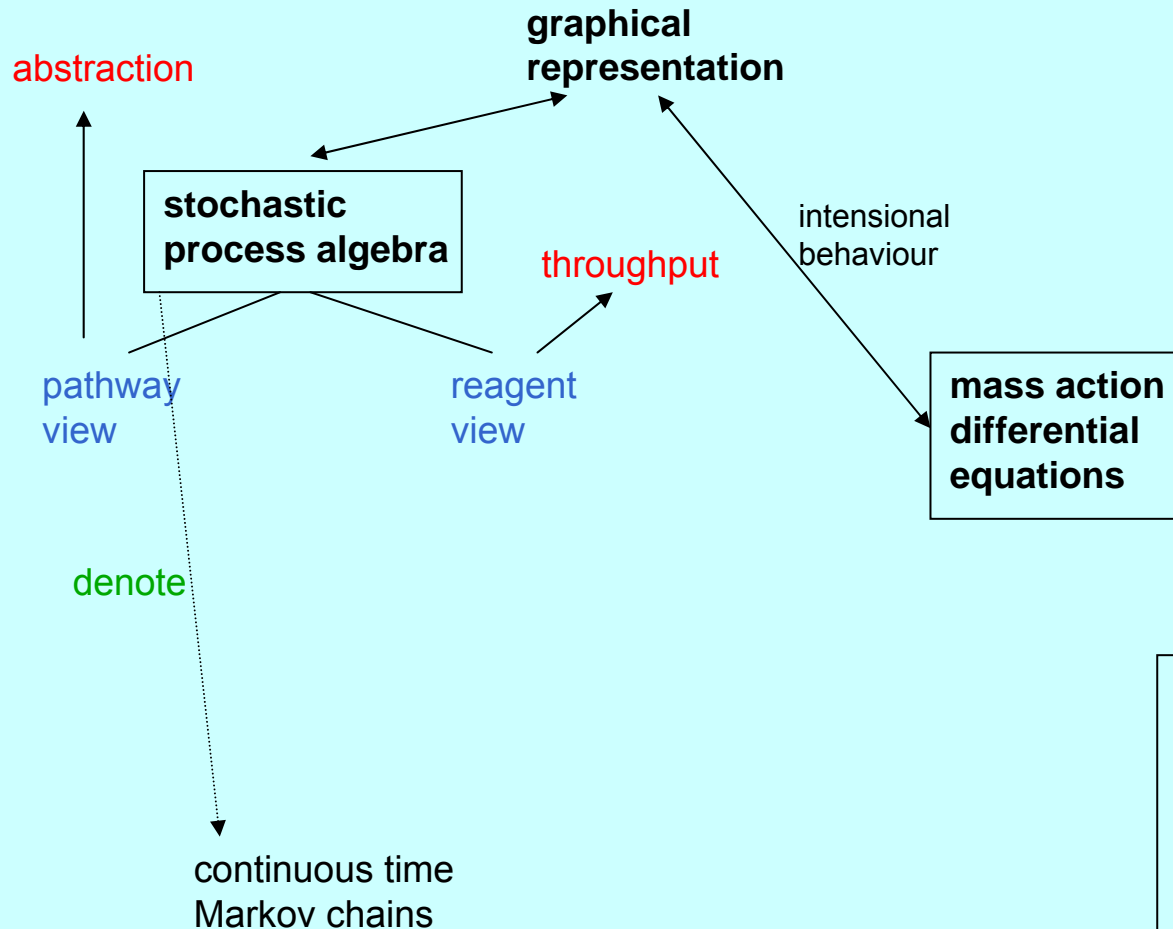
With simple formulae (*rewards*) it is possible to reproduce a simulation trace of the system. Compare with ODE model:



Levels	ϵ_a	ϵ_r	$C\epsilon_a$	$C\epsilon_a^2$
4	0.126 mM	0.280	21.557 mM	2.58
5	0.103 mM	0.217	17.569 mM	1.727
6	0.086 mM	0.176	14.582 mM	1.191
8	0.061 mM	0.122	10.402 mM	0.605
12	0.036 mM	0.071	6.042 mM	0.204

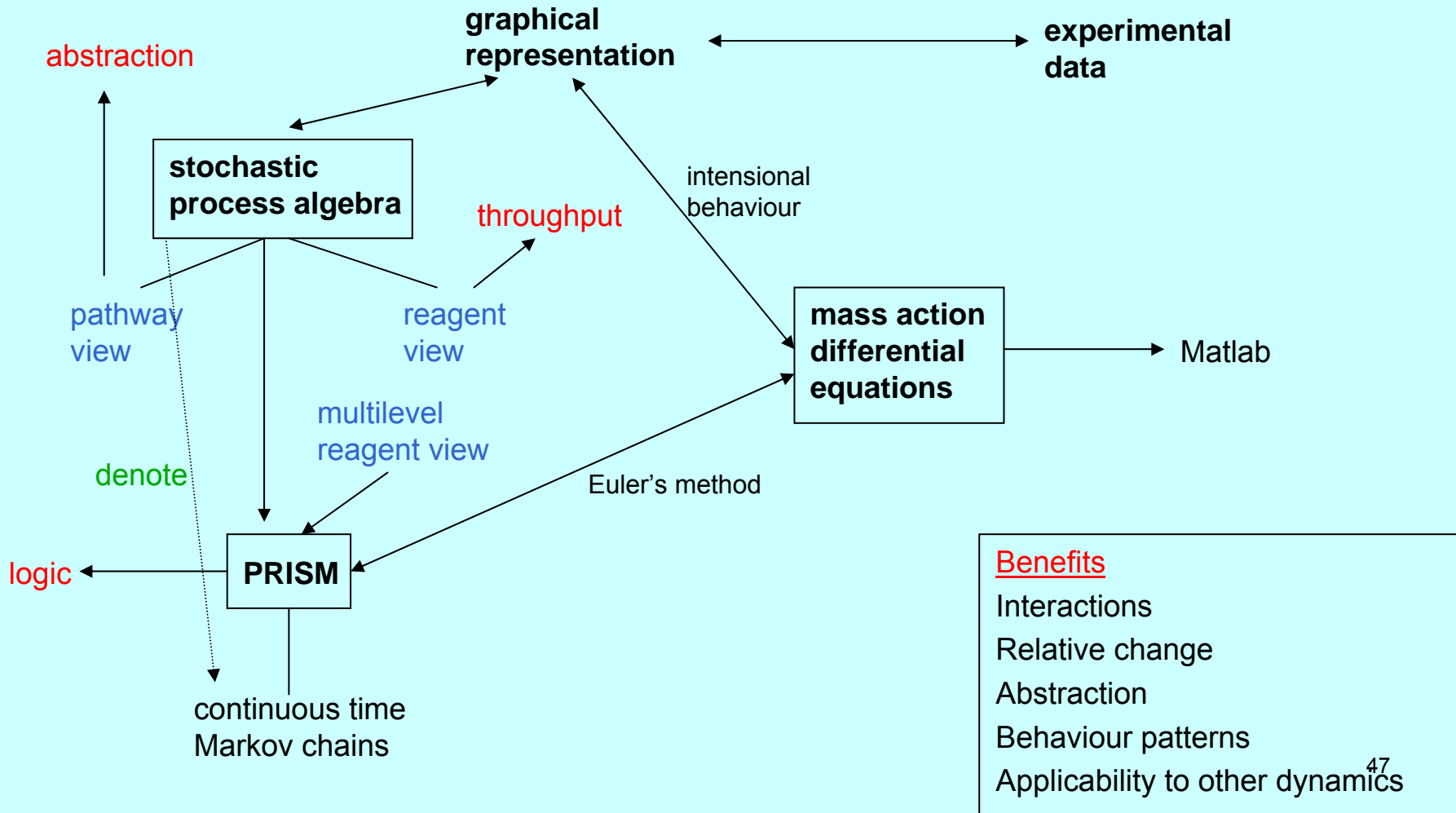
7-9 levels are sufficient! Extremely tractable!

Big Picture



- Benefits
- Interactions
 - Relative change
 - Abstraction
 - Behaviour patterns
 - Applicability to other dynamics⁴⁶

Bigger Picture



Modelling

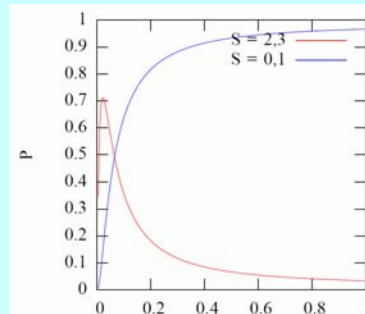
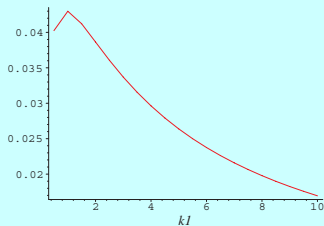
Each process molecular *species* is modelled by a stochastic, concurrent process

Expressed using high level languages from computer science!

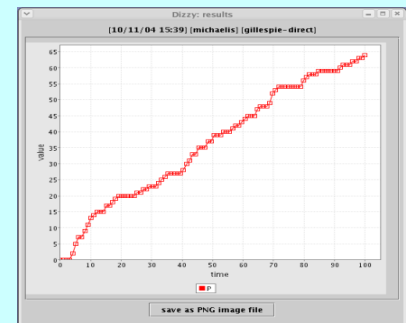
⇒ new kinds of analysis
new ways of relating traditional and non-standard models

Process algebra models (reagent and pathway)

Throughput of $k\delta$ product



$$\frac{dm_3}{dt} = k_1 m_1 m_2$$



$$S = ?[(Raf-1^* \geq 2) \wedge Raf-1^* \leq 3]$$

The really big picture

topology & parameters + stoichiometry + traffic + data

$\begin{Bmatrix} 00011100 \\ 00110000 \\ 00010000 \end{Bmatrix}$

rates

stochastic process algebra

Raf-1*_H = (k1react,k1). Raf-1*_L + ...

stochastic simulation
Dizzy

Approximation
model-fitting
(Bayesian inference)

abstraction

throughput

approximation

pathway
view

reagent
view

Continuous time
Markov chains

N levels

solve

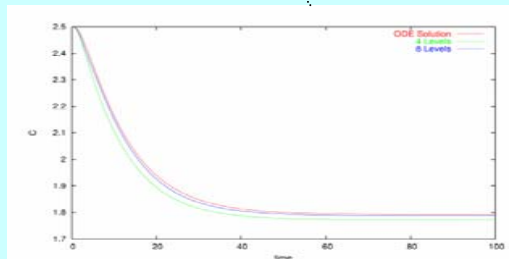
continuous
stochastic
logic

probabilities

PRISM

approximation

**mass action
differential
equations**

$$\frac{dm1}{dt} = -k1*m1*m2 + k2*m3$$


simulation

(other dynamics)

continuous time
discrete concentrations

continuous time
continuous concentrations

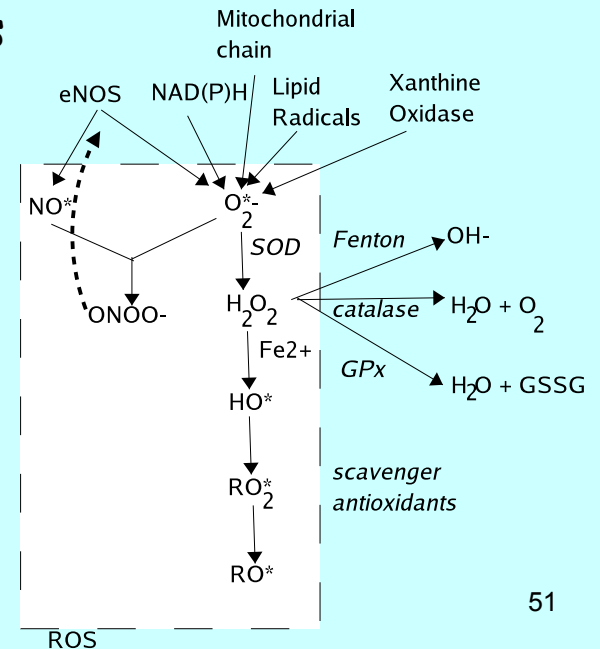
discrete time
continuous concentrations

Discussion & Conclusions

- Regent-centric view
 - probabilities of states (H/L)
 - differential equations
 - fit with data
 - PEPA \rightarrow PRISM
- Pathway-centric view
 - simpler model
 - building blocks, modularity approach
 - no further information is gained from having multiple levels
- Life science
 - see potential of an *interaction* approach which affords two views; static analysis and modelling language
- Computing science
 - individual/population view
 - continuous, traditional mathematics and simulation
 - theorem: as $n \rightarrow \infty$, ODE and CTMC semantics converge (1st, 2nd order eqns)

Further Challenges

- Quantification of abstraction over networks
 - “chop” off bits of network
- Model spatial dynamics (vesicles) and scaffolds
- Model other stoichiometries
- Relate individual to molar concentration models
- Prove CTMC and ODE equivalence
- Relate data to high level operators (topology)
- Further applications: e.g. oxidative stress cardiovascular medicine



The End

Thank you.